Nutritional and Antioxidant Values of Oyster Mushroom (P. Sajor-caju) Cultivated on Rubber Sawdust

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Abstract—Grey oyster mushroom (Pleurotus sajor-caju) is one of the edible mushrooms from Pleurotus family, which is cultivated commercially in Malaysia. The global mushroom market has shown remarkable growth in recent years and has attractive market potential for the future. This study aimed to determine nutritional value and antioxidant properties of fresh grey oyster mushroom (Pleurotus sajor-caju) with the intention to give better information on nutritional composition of fresh oyster mushroom cultivated on rubber sawdust. Grey oyster mushrooms were freshly harvested from Teloi Agro Farm, Kedah, Malaysia. The proximate analysis was used to determine nutrient composition of grey oyster mushroom. Highest nutritional value of grey oyster mushroom is the moisture content (90.10±0.09%), followed by crude fibre content (17.27±1.08%), protein content (4.00±0.31%), carbohydrates content (3.54±0.09%), while the lowest nutritional value was possessed by fat content (1.18±0.33%) and ash content (1.16±0.02%). The antioxidant activities of the mushroom were measure using total phenolic content (TPC) by Folin-Ciocalteu reagent method and 2, 2-diphenyl-1-picrylhydrazyl radical scavenging (DPPH) assay. The hot water extract showed mushroom powder possessed 2.21±0.03 mg GAE/g of dried mushroom powder for total phenolic content and at concentration of 0.1g/ml it can inhibit 89.29% of DPPH radicals. By virtue of having high fibre with low fat and high antioxidant activity, grey oyster mushroom can be considered as a functional food, which can provide health benefits.

Keywords—P. sajor-caju; grey oyster mushroom; sawdust; antioxidant properties; nutritional value

I. INTRODUCTION

Mushroom was said to be first cultivated in China as early as 600AD [1]. Consumption of mushrooms is growing fast due to its balanced nutritional composition [2]-[4] and will give benefits for health improvement because they are low in calories, sodium, fat and cholesterol but rich in fibre, vitamins and minerals [5]. More than that, oyster mushrooms is capable of handling the problem of protein malnutrition in developing and underdeveloped countries [6].

Grey oyster mushroom is one of the 17 mushroom varieties which can be grown in Malaysia [7]. Their growth is suitable with Asian climate, they can be grown in simple and cheaper production costs [8] make them easily available all around the year. Grey oyster mushroom can be cultivated on wide selection of substrates such as rubber sawdust, paddy straw, wood log and sugar baggase which contain high amount of lignocellulosic material [1]. The total demand for mushrooms in Malaysia is expected to increase from 23, 000 tons per year (2008) to 72, 000 tons per year in 2020 [9].

P. sajor-caju also known by its common name as grey oyster mushroom, and this name was obtained for the physical appearance which resembled an oyster shell. It was first isolated from India [10],[11] this strain has been identified to be the best strain for cultivation in tropical and subtropical regions [10]-[13]. In Malaysia, oyster mushroom are usually consumed as soup, grilled or deep fried and it has a great demand as local street food.

Among the three species of mushrooms commercially cultivated in India (Agaricus bisporus, Volvariella volvacea, Pleurotus sajor-caju), P. sajor-caju was said to possess unique nutritional and medicinal values, aroma as well as taste [5]. Antioxidants present in mushrooms are possible to be protective agents to help human body to reduce oxidative damage without any interference [14]. [15] agreed that consumption of plant foods can provides protection against various diseases including cancer and cardio- and cerebrovascular diseases.

In this research, proximate composition and antioxidant properties of P. sajor-caju was evaluated to give some contribution on nutritional values information of P.sajor-caju cultivated on rubber sawdust in Malaysia.

II. MATERIALS AND METHODS

Kjeldahl Catalyst Tablet was purchased from Merck (Germany). Whereas 2,2-Diphenyl-1-picrylhydrazyl (DPPH), Folin & Ciocalteu’s phenol reagent was obtained from...
Sigma-Aldrich (St. Louis, MO., USA). Ascorbic acid and gallic acid were procured from R&M (Essex, UK). Petroleum ether (40 – 60 °C) was purchased from Fisher Scientific (Dublin, Ireland). All other chemicals and reagents used were analytical grade.

A. Samples
Oyster mushroom was freshly purchased from Teloi Agro Farm Enterprise, Kedah, Malaysia in the early morning. Sample was stored in ice box to retain its freshness before transported for analysis in the USM.

B. Proximate composition
Proximate composition was done on moisture, fat, crude fibre and ash [16]. Protein factor used in determining crude protein is 4.8 [17], [18] while carbohydrate was calculated by difference.

C. Extraction
Antioxidant extraction was carried out according to [19] with slight modification. 2 g of ground samples were mixed into 50 ml of hot water and stirred for 15 minutes. Samples were centrifuged at 2000g for 15 minutes. Supernatants were collected for analysis.

D. Radical scavenging activity (RSA) on DPPH
RSA was determined by using a method proposed by [20]. 0.1 ml of samples extraction was mixed with 2.9 ml of 100µM methanolic DPPH (2,2-diphenyl-1-picrylhydrazyl) and vortex mixed for a minute. Absorbance was read immediately at 517 nm wavelength with Shimadzu UV-vis spectrophotometry, Japan. Vitamin C (ascorbic acid) was used as standard. Radical scavenging activity (RSA) was calculated using the formula:

\[
\% \text{ RSA} = \frac{(A_o - A_s)}{A_o} \times 100
\]

Where \(A_o\) is the absorbance of 100 µM methanolic DPPH only and \(A_s\) is the absorbance of reaction mixture.

E. Total phenolic content
Folin-phenol method [19] was used to determine total phenolic content in mushroom samples. 100µL of sample aliquot was added into 900µL of distilled water. 1 ml of 1N Folin-Ciocalteu’s Reagent and 2 ml of 10% sodium carbonate solution were added and mixed before they were incubated for 1 hour at room temperature. Absorbance was measured at 765 nm with Shimadzu UV-visible spectrophotometer, Japan. Vitamin C (ascorbic acid) was used as standard. Total phenolic content (TPC) was determined by using the formula:

\[
\text{TPC} = \frac{150 \times \text{Absorbance}}{\text{Concentration of gallic acid}}
\]

A. NF= protein factor 4.38 \(D = \) Calculated by difference.

Table 1: Proximate Composition of Fresh P. sajor-caju Cultivated on Rubber Sawdust.

<table>
<thead>
<tr>
<th>Nutritional Composition</th>
<th>% Fresh Weight</th>
</tr>
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<tbody>
<tr>
<td>Ash</td>
<td>1.16±0.02</td>
</tr>
<tr>
<td>Moisture</td>
<td>90.10±0.09</td>
</tr>
<tr>
<td>Protein</td>
<td>4.00±0.31</td>
</tr>
<tr>
<td>Fat</td>
<td>1.18±0.33</td>
</tr>
<tr>
<td>Crude Fiber</td>
<td>17.27±1.08</td>
</tr>
<tr>
<td>Carbohydrate</td>
<td>3.54±0.09</td>
</tr>
</tbody>
</table>

III. RESULTS AND DISCUSSIONS
The results of nutritional value obtained for the study are shown in Table 1. Ash content is recorded at 1.16±0.02% whereas moisture is 90.10±0.09%. Ash content signifies mineral content in mushrooms as has been stated by [21] which was possibly resulted from potassium [22]. Phosphorus, magnesium [23], calcium, copper, iron and zinc [24], [25] has studied minerals content of P. sajor-caju were varied if cultivated on different substrate.

P. sajor-caju has proven to contains high water percentage when incorporated into chicken patty, it helped to reduce the hardness of chicken patty compared to the non-incorporated one [26]. This high percentage of water is depended on several factors such as watering frequency, weather and storage condition thus making them highly perishable vegetables in the market.
whereas non-digestible form makes up the large component of total carbohydrates of mushrooms [27].

The DPPH method was used to determine the free radical scavenging activity of Pleurotus sajor-caju. The radical scavenging activity (RSA) of mushroom extracts was tested against the DPPH. The % RSA for P. sajor-caju are presented in Fig. 1. Antioxidant compounds exist in the sample will transfer electron or hydrogen atom to react with DPPH. The odd electron in the DPPH free radical gives a strong absorption maximum at 517 nm and is purple in color [31]. Reacted DPPH will turn the color from purple to yellow. RSA of mushroom sample varied from 64.51-89.29% (fig. 1) and increased with the increase in concentration. At 100% concentration, it can inhibit 89.29% DPPH. A study done by [32] proved that P. sajor-caju showed comparatively lower RSA than P. eryngii, P. djamor and P. flabellatus (67.4–69.67% at 100% mycelial extract concentration). However, in this study, the RSA of P. sajor-caju is determined to be at higher concentration.

Total phenolic content of mushroom extraction was estimated by Folin-Ciocalteu method and results were expressed as mg of gallic acid equivalents per gram of mushroom powder. Total phenolic contents was determined at 2.21±0.03mg/g in this study as compared to the other type of mushrooms species such as Agaricus bisporus (0.63 mg/g), Hypocissus marmorosus (0.67 mg/g), Volvariella volvacea (0.73 mg/g), Flammulina velutipes (0.75 mg/g), Pleurotus eryngii (0.44 mg/g), Pleurotus ostreatus (0.39 mg/g), Lentinula edodes (0.49 mg/g), Hericium erinaceus (0.46 mg/g) [33]. These phenolic compounds could be used as an important indicator of antioxidant capacity [32]. Higher antioxidant activity could be exhibited with the presence of the greater numbers of hydroxyl groups in the phenolics [34]. Consumption of Pleurotus sajor-caju can be beneficial for protecting body from harmful free-radicals compounds which can create oxidative damage to the body.

IV. CONCLUSIONS

Analyses of the proximate composition of commonly cultivated Pleurotus sajor-caju reveal that they are rich in fibre with low fat and high antioxidant activity. Grey oyster mushroom can be considered as a functional food, which can provide health benefits.


