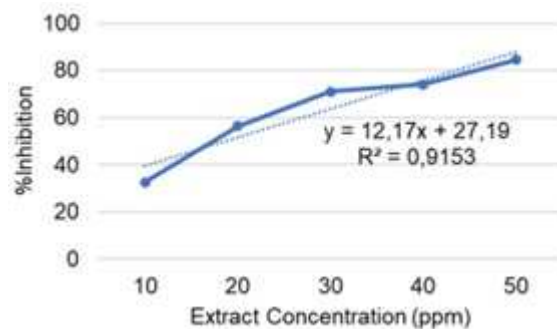
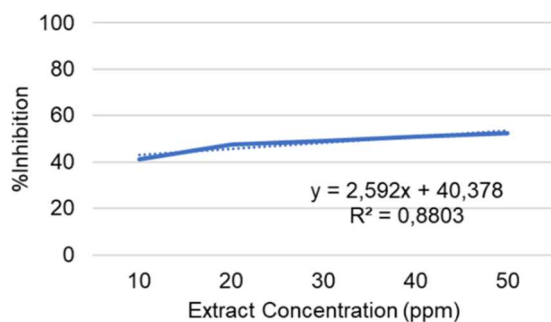


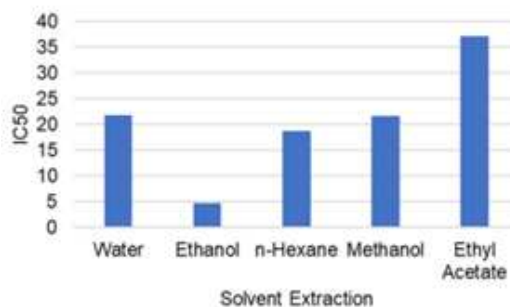
(c)



(d)



(e)



(f)

Fig. 1 The DPPH inhibitory and IC<sub>50</sub> of Gayo Arabica Coffee Cascara Pulp Extract with varied solvent extractions, A. water extract, B. ethanol extract, C. methanol extract, D. n-hexane extract, E. ethyl acetate extract, F. IC<sub>50</sub> revealing antioxidant activity

### B. Secondary metabolites profile of cascara Gayo arabica coffee

Various solvents for extraction performed varied antioxidant activities, and varied solvents also affected the metabolites profile. Table 1—3 shows there are several solvents with the same compounds, ethanol, methanol, and ethyl acetate, with 5-Hydroxymethylfurfural compounds that function as antioxidants, antibacterial, and antiproliferative. However, the concentrations of the compounds in the three solvents were different. The highest concentration was found in ethanol solvent, and the lowest concentration of 5-Hydroxymethylfurfural was found in ethyl acetate solvent.

TABLE I  
METABOLITE PROFILES OF CASCARA GAYO ARABICA COFFEE IN ETHANOL SOLVENT

Retention time	% Area	Compound	Biological Activity
21.377	22.31	5-Hydroxymethylfurfural	Antioxidant, antibacterial, and antiproliferative [32]
29.277	21.07	Caffeine	Antioxidant and antimicrobial [33]
30.701	12.85	n-Hexadecanoic acid	Anti-cancer, antioxidant, anti-cancer, anti-alopecic, anti-inflammatory [34] 35]
32.435	11.94	Cis-Vaccenic acid	Antibacterial, hypolipidemic effect, antioxidant, anti-inflammatory [36]
28.005	3.93	Hydrazinecarboxamide, 2-(2-methylcyclohe)	Anticonvulsant [37]

TABLE II  
METABOLITE PROFILES OF CASCARA GAYO ARABICA COFFEE IN METHANOL SOLVENT

Retention time	% Area	Compound	Biological Activity
21.830	16.61	5-Hydroxymethylfurfural	Antioxidant, antibacterial, and antiproliferative [32]
29.326	12.55	Caffeine	Antioxidant and antimicrobial [33]
28.295	10.59	Quinic Acid	Anti-HIV-1 Antioxidant [38] 37]
30.706	7.38	n-Hexadecanoic acid	Anti-cancer, antioxidant, anti-cancer, anti-alopecic, and anti-inflammatory [34] 35]
32.416	5.71	cis-Vaccenic acid	, antibacterial, hypolipidemic effect, antioxidant, and anti-inflammatory [36]

TABLE III  
METABOLITE PROFILES OF CASCARA GAYO ARABICA COFFEE IN N-HEXANE SOLVENT

Retention time	% Area	Compound	Biological Activity
28.934	57.53	Caffeine	Antioxidant and antimicrobial [33]
35.413	5.94	Hexadecanoic acid, 2-hydroxy-1-(hydroxyl)	No activity was reported [40]
30.534	5.26	Palmitic acid n-Hexadecanoic acid	Anti-cancer antioxidant, anti-cancer, anti alopecic, anti-inflammatory [34], [35]
36.947	4.50	9-Octadecenoic acid (Z)-, 2,3-dihydroxypropyl ester	Hepatoprotective activity [41], anti-cancer [42], antifungal, antibacterial [43]
30.029	4.34	Hexadecanoic acid, methyl ester (CAS) methyl palmitate	Antioxidant, hemolytic, hypocholesterolemic [43]

In addition, the n-hexadecanoic acid (Table 4) compound, which functions as an antioxidant, is also found in several

solvents, namely ethanol and n-hexane solvents, but with different compound concentrations, the highest concentration of n-hexadecenoic acid compounds is found in ethanol solvents. The caffeine compound that functions as an antioxidant and antimicrobial is also found in several solvents, namely ethanol, methanol, n-Hexane, and ethyl acetate, with caffeine compound concentration found in n-Hexane solvent, 57.53%, and the lowest caffeine concentration was found in methanol solvent, 12.55%.

The water and methanol solvent fractions also have the same compound, quinic acid, which functions as an antioxidant with the highest concentration in the water solvent fraction. The utilization of natural products in the cosmetic application is becoming a great interest nowadays as phenolic compound biological activity proved to have anti-aging activity and the ability to protect skin from ultraviolet radiation [31]. An animal study conducted by Marija et al. [47] shows that a dietary supplement derived from a vegetable from the genus Brassica reduced the effect of skin aging. Puxvadee et al. [48] use *Phyllanthus emblica L.* branch in topical gel to evaluate the efficacy of the anti-aging activity. Clinical improvement was observed in the effect of skin aging, such as reduction of skin wrinkles, enhanced skin elasticity, and lightening skin color.

TABLE IV  
METABOLITE PROFILES OF CASCARA GAYO ARABICA COFFEE IN ETHYL ACETATE SOLVENT

Retention time	% Area	Compound	Biological Activity
21.187	13.88	5-Hydroxymethylfurfural	Antioxidant, antibacterial and antiproliferative [32]
29.130	13.81	Caffeine	Antioxidant and antimicrobial [33]
27.307	9.83	1(2H)-Naphthalenone, octahydro-4a,8a-dim	Antioxidant [44]
30.605	8.16	n-Hexadecanoic acid	Anti-cancer, antioxidant, antiallopecic, anti-inflammatory [34],[ 35]
32.347	6.90	Oleic acid	Lower blood pressure, anti-cancer, hepatoprotective, anti-inflammatory [45], [46]

#### IV. CONCLUSION

In conclusion, various solvents affected the antioxidant and metabolite profiles of cascara Gayo Arabica coffee extracts. The ethanolic extract performs the high antioxidant activity with the lowest IC50 of DPPH inhibition, while the lowest antioxidant activity was ethyl acetate solvent treatment. Polar and non-polar solvents performed different metabolites and shifted the retention time of compounds.

#### ACKNOWLEDGMENT

The authors thank Syiah Kuala University for all the knowledge and support in every research process and information for the study.

#### REFERENCES

- [1] N. Juliaviani, Sahara, and R. Winandi, "Transmisi Harga Kopi Arabika Gayo Di Provinsi Aceh," *Jurnal Agribisnis Indonesia*, vol. 5, no. 1, pp. 39–56, 2017.
- [2] S. Mulato, E. Suharyanto, and Kaswanto., "Products Development of Coffee. Indonesian Coffee and Cocoa," *Research Institute Jember*, vol. 9, no. 4, pp. 1–8, 2020.
- [3] A. Iriondo-dehond, M. Iriondo-dehond, and M. Castilo "Applications of Compounds from Coffee Processing By-Products," *Biomolecules*, vol. 10, no. 1219, 2020. doi:10.3390/biom10091219
- [4] J. Sangta *et al.*, "Recovery of polyphenolic fraction from arabica coffee pulp and its antifungal applications," *Plants*, vol. 10, no. 7, pp. 1–15, 2021, doi: 10.3390/plants10071422.
- [5] F. G. Gemechu, "Embracing nutritional qualities, biological activities and technological properties of coffee byproducts in functional food formulation," *Trends Food Sci Technol*, vol. 104, pp. 235–261, 2020, doi:10.1016/j.tifs.2020.08.005.
- [6] P. Esquivel and V. M. Jiménez, "Functional Properties of Coffee and Coffee By-products," *Food Research International*, vol. 5, no. 3, pp. 1–9, 2012, doi: 10.1016/j.foodres.2011.05.028.
- [7] Y. Yang and T. Zhang, "Antimicrobial activities of tea polyphenol on phytopathogens: A review," *Molecules*, vol. 24, no. 4, 2019, doi: 10.3390/molecules24040816.
- [8] Y. Fukushima *et al.*, "Consumption of polyphenols in coffee and green tea alleviates skin photoaging in healthy Japanese women," *Clin Cosmet Investig Dermatol*, vol. 13, pp. 165–172, 2020, doi: 10.2147/CCID.S225043.
- [9] S. Soleymani, M. H. Farzaei, A. Zargaran, S. Niknam, and R. Rahimi, "Promising plant-derived secondary metabolites for treatment of acne vulgaris: a mechanistic review," *Arch Dermatol Res*, vol. 312, no. 1, pp. 5–23, 2020, doi: 10.1007/s00403-019-01968-z.
- [10] A. Heeger, A. Kosińska-Cagnazzo, E. Cantergiani, and W. Andlauer, "Bioactives of coffee cherry pulp and its utilisation for production of Cascara beverage," *Food Chem*, vol. 221, pp. 969–975, Apr. 2017, doi: 10.1016/J.FOODCHEM.2016.11.067.
- [11] K. Hasballah, W. Lestari, M. Y. Listiawan, and S. Sofia, "Coffee by-products as the source of antioxidants: A systematic review," *F1000Res*, vol. 11, pp. 1–12, 2022, doi: 10.12688/f1000research.107811.1.
- [12] P. F. Surai, I. I. Kochish, V. I. Fisinin, and M. T. Kidd, "Antioxidant defence systems and oxidative stress in poultry biology: An update," *Antioxidants*, vol. 8, no. 7, pp. 1–36, 2019, doi: 10.3390/antiox8070235.
- [13] X.-J. Shen *et al.*, "Progress in Phytochemical and Bioactivities of Coffea arabica L.," *Medicine Research*, vol. 4, no. 3, p. 200012, 2020, doi: 10.21127/yaoyimr20200012.
- [14] J. W. Shin *et al.*, "Molecular mechanisms of dermal aging and anti-aging approaches," *Int J Mol Sci*, vol. 20, no. 9, 2019, doi: 10.3390/ijms20092126.
- [15] M. C. McCabe *et al.*, "Alterations in extracellular matrix composition during aging and photoaging of the skin," *Matrix Biol Plus*, vol. 8, Nov. 2020, doi: 10.1016/j.mbplus.2020.100041.
- [16] E. Hanani, *Analisis Fitokimia*. Jakarta: EGC, 2015.
- [17] M. Insanu, I. Fidrianny, N. H. H. Imtinan, and S. Kusmardiyani, "Liberica coffee (Coffea liberica L.) from three different regions: In vitro antioxidant activities," *Biointerface Res Appl Chem*, vol. 11, no. 5, pp. 13031–13041, 2021, doi: 10.33263/BRIAC115.1303113041.
- [18] J. W. Honour, "Gas Chromatography-Mass Spectrometry BT - Hormone Assays in Biological Fluids," M. J. Wheeler and J. S. M. Hutchinson, Eds. Totowa, NJ: Humana Press, 2006, pp. 53–74. doi: 10.1385/1-59259-986-9:53.
- [19] De-hond AI and A. Elizondo, "Assessment of Healthy and Harmful Maillard Reaction Products in a Novel Coffee Cascara Beverage.," *Journal of Food and Nutrition Research*, vol. 620, no. 9, pp. 1–18, 2020.
- [20] A. Pua *et al.*, "A systematic study of key odourants, non-volatile compounds, and antioxidant capacity of cascara (dried Coffea arabica pulp)," *Lwt*, vol. 138, p. 110630, 2021, doi: 10.1016/j.lwt.2020.110630.

- [21] A. Hejna, "Potential applications of by-products from the coffee industry in polymer technology – Current state and perspectives," *Waste Management*, vol. 121, pp. 296–330, 2021, doi: 10.1016/j.wasman.2020.12.018.
- [22] A. Ontawong *et al.*, "Lipid-lowering effects of Coffea arabica pulp aqueous extract in Caco-2 cells and hypercholesterolemic rats," *Phytomedicine*, vol. 52, pp. 187–197, 2019, doi: 10.1016/j.phymed.2018.06.021.
- [23] E. Sholichah, D. Desnilasari, R. Subekti, M. A. Karim, and B. Purwono, "The influence of coffee cherry fermentation on the properties of Cascara arabica from Subang, West Java The," *IOP Conf. Series: Materials Science and Engineering*, vol. 1011, 2021, doi: 10.1088/1757-899X/1011/1/012006.
- [24] N. Arpi, M. Muzaifa, M. I. Sulaiman, R. Andini, and S. I. Kesuma, "Chemical Characteristics of Cascara, Coffee Cherry Tea, Made of Various Coffee Pulp Treatments," *IOP Conf. Series: Earth and Environmental Science*, vol. 709, 2021, doi: 10.1088/1755-1315/709/1/012030.
- [25] G. G. Marcheafave, C. D. Tormena, E. D. Pauli, M. Rakocevic, R. E. Bruns, and I. S. Scarminio, "Experimental mixture design solvent effects on pigment extraction and antioxidant activity from Coffea arabica L. leaves," *Microchemical Journal*, vol. 146, pp. 713–721, 2019, doi:10.1016/j.microc.2019.01.073.
- [26] O. R. Alara, N. H. Abdurahman, and C. I. Ukaegbu, "Extraction of phenolic compounds: A review," *Curr Res Food Sci*, vol. 4, pp. 200–214, 2021, doi:10.1016/j.crfss.2021.03.011.
- [27] N. M. Peixoto Araujo *et al.*, "Enzymatic treatment improves the antioxidant and antiproliferative activities of Adenanthera pavonina L. seeds," *Biocatal Agric Biotechnol*, vol. 18, p. 101002, 2019, doi: 10.1016/j.bcab.2019.01.040.
- [28] H. D. F. Q. Barros, A. M. Baseggio, C. F. F. Angolini, G. M. Pastore, C. B. B. Cazarin, and M. R. Marostica-Junior, "Influence of different types of acids and pH in the recovery of bioactive compounds in Jabuticaba peel (Plinia cauliflora)," *Food Research International*, vol. 124, pp. 16–26, 2019, doi:10.1016/j.foodres.2019.01.010.
- [29] K. Kiattisin, N. Intasai, N. Nitthikan, and T. Nantararat, "Antioxidant, Anti-tyrosinase, Anti-aging Potentials and Safety of Arabica Coffee Cherry Extract," *Chiang Mai Journal of Science*, vol. 46, no. 5, pp. 930–945, 2019.
- [30] M. S. Alkaltham, A. Salamatullah, and K. Hayat, "Determination of coffee fruit antioxidants cultivated in Saudi Arabia under different drying conditions," *Journal of Food Measurement and Characterization*, vol. 14, no. 3, pp. 1306–1313, 2020, doi: 10.1007/s11694-020-00378-4.
- [31] D. J. de Lima Cherubim, C. V. Buzanello Martins, L. Oliveira Fariña, and R. A. da Silva de Lucca, "Polyphenols as natural antioxidants in cosmetics applications," *J Cosmet Dermatol*, vol. 19, no. 1, pp. 33–37, 2020, doi: 10.1111/jocd.13093.
- [32] M. T. Yassin, A. A. Mostafa, and A. A. Al Askar, "In Vitro Evaluation of Biological Activities and Phytochemical Analysis of Different Solvent Extracts of Punica granatum L. (Pomegranate) Peels," *Plants*, vol. 10, no. 12, 2021. doi: 10.3390/plants10122742.
- [33] W. Khochapong, S. Ketnawa, Y. Ogawa, and N. Punbusayakul, "Effect of in vitro digestion on bioactive compounds, antioxidant and antimicrobial activities of coffee (Coffea arabica L.) pulp aqueous extract," *Food Chem*, vol. 348, p. 129094, 2021, doi: 10.1016/j.foodchem.2021.129094.
- [34] B. Bharath, K. Perinbam, S. Devanesan, M. S. AlSalhi, and M. Saravanan, "Evaluation of the anti-cancer potential of Hexadecanoic acid from brown algae Turbinaria ornata on HT–29 colon cancer cells," *J Mol Struct*, vol. 1235, p. 130229, 2021, doi: 10.1016/j.molstruc.2021.130229.
- [35] A. Nepal, M. Chakraborty, D. Sarma, and P. K. Haldar, "Phytochemical Characterization of Aeschynanthus sikkimensis (Clarke) Stapf. (Gesneriaceae) using GC-MS," *International Journal of Pharmaceutical Research*, vol. 13, no. 3, 2021, doi: 10.31838/ijpr/2021.13.03.053.
- [36] O. Ajanaku Christiana, O. Echeme Johnbull, C. Mordi Raphael, O. Olugbuyiro Joseph, M. Osamudiamen Paul, and G. Jolayemi Emmanuel, "Gas Chromatographic Study of Bio-active Compounds in Methanolic Extract of Leaf of Crateva adansonii DC," in *Journal of Physics: Conference Series*, Oct. 2019, vol. 1299, no. 1. doi: 10.1088/1742-6596/1299/1/012014.
- [37] A. Oluwaseye, A. Uzairu, G. A. Shallangwa, and S. E. Abechi, "A novel QSAR model for designing, evaluating, and predicting the anti-MES activity of new 1H-pyrazole-5-carboxylic acid derivatives," *Journal of the Turkish Chemical Society, Section A: Chemistry*, pp. 739–774, Jul. 2017, doi: 10.18596/jotcsa.304584.
- [38] S. E. Yazdi, G. Prinsloo, H. M. Heyman, C. B. Oosthuizen, T. Klimkait, and J. J. M. Meyer, "Anti-HIV-1 activity of quinic acid isolated from Helichrysum mimetes using NMR-based metabolomics and computational analysis," *South African Journal of Botany*, vol. 126, pp. 328–339, 2019, doi: https://doi.org/10.1016/j.sajb.2019.04.023.
- [39] M. Karaman *et al.*, "Polarography as a technique of choice for the evaluation of total antioxidant activity: The case study of selected Coprinus Comatus extracts and quinic acid, their anti-diabetic ingredient," *Nat Prod Res*, vol. 35, no. 10, pp. 1711–1716, May 2021, doi: 10.1080/14786419.2019.1628753.
- [40] K. Ravichandiran and M. Parani, "Comparative GC-MS Analysis of Methanolic Extracts from Different Parts of Bitter Gourd (Momordica charantia L.) Fruit," *Journal of Food and Nutrition Research*, vol. 9, pp. 102–107, Mar. 2021, doi: 10.12691/jfnr-9-3-1.
- [41] S. M. Osman, A. E. El-Haddad, M. A. El-Raey, S. M. A. El-Khalik, M. A. Koheil, and M. Wink, "A new octadecenoic acid derivative from caesalpinia gilliesii flowers with potent hepatoprotective activity," *Pharmacogn Mag*, vol. 12, no. 46, pp. S332–S336, 2016, doi: 10.4103/0973-1296.185752.
- [42] M. Simonovic, V. Kojic, D. Jakimov, M. Glumac, and B. Pejin, "Raspberry seeds extract selectively inhibits the growth of human lung cancer cells in vitro.," *Nat Prod Res*, vol. 35, no. 13, pp. 2253–2256, 2021. doi: 10.1080/14786419.2019.1666391.
- [43] S. Arora and G. Kumar, "Phytochemical screening of root, stem and leaves of Cenchrus biflorus Roxb.," *Journal of Pharmacognosy and Phytochemistry*, vol. 7, no. 1, pp. 1445–1450, 2018.
- [44] E. A. Sunday, N. C. Nnedimma, W. G. Peter, and G. B. Orlando, "Screening for Pharmacological Compounds and Antioxidant Activity of Hedychium coronarium J. Koenig," *Asian Journal of Research in Botany*, vol. 5, no. 4, pp. 34–47, 2021.
- [45] A. M. Tindall, C. J. McLimans, K. S. Petersen, P. M. Kris-Etherton, and R. Lamendella, "Walnuts and Vegetable Oils Containing Oleic Acid Differentially Affect the Gut Microbiota and Associations with Cardiovascular Risk Factors: Follow-up of a Randomized, Controlled, Feeding Trial in Adults at Risk for Cardiovascular Disease," *Journal of Nutrition*, vol. 150, no. 4, pp. 806–817, Apr. 2020, doi: 10.1093/jn/nxz289.
- [46] E. Piccinin *et al.*, "Role of oleic acid in the gut-liver axis: From diet to the regulation of its synthesis via Stearoyl-CoA desaturase 1 (SCD1)," *Nutrients*, vol. 11, no. 10. MDPI AG, Oct. 01, 2019. doi: 10.3390/nu11102283.
- [47] M. Petkovic *et al.*, "Dietary supplementation with sulforaphane ameliorates skin aging through activation of the Keap1-Nrf2 pathway," *J Nutr Biochem*, vol. 98, p. 108817, 2021, doi: 10.1016/j.jnutbio.2021.108817.
- [48] P. Chaikul, M. Kanlayavattanukul, J. Somkumnerd, and N. Lourith, "Phyllanthus emblica L. (amla) branch: A safe and effective ingredient against skin aging," *J Tradit Complement Med*, vol. 11, no. 5, pp. 390–399, 2021, doi: 10.1016/j.jtcm.2021.02.004.